

## Review

## Buried Treasure: Evolutionary Perspectives on Microbial Iron Piracy

Matthew F. Barber<sup>1,\*</sup> and Nels C. Elde<sup>1</sup>

**Host-pathogen interactions provide valuable systems for the study of evolutionary genetics and natural selection. The sequestration of essential iron has emerged as a crucial innate defense system termed nutritional immunity, leading pathogens to evolve mechanisms of 'iron piracy' to scavenge this metal from host proteins. This battle for iron carries numerous consequences not only for host-pathogen evolution but also microbial community interactions. Here we highlight recent and potential future areas of investigation on the evolutionary implications of microbial iron piracy in relation to molecular arms races, host range, competition, and virulence. Applying evolutionary genetic approaches to the study of microbial iron acquisition could also provide new inroads for understanding and combating infectious disease.**

### An Evolving View of Host–Microbe Interactions

The outcome of an infection can have profound consequences for both host and pathogen populations. Intense selective pressures make host–pathogen interactions an attractive biological model to study evolutionary genetics over relatively short intervals of time. To date, much work has focused on rapid evolution involving canonical host immune defenses or antibiotic resistance [1,2]. However, we now know that hosts possess numerous additional means to restrict pathogens, including factors engaged in other core physiological functions. Nutrient iron sequestration provides one such alternative mode of host defense against bacteria and eukaryotic pathogens [3]. Iron is an essential micronutrient for microbes, as well as their hosts, due to its ability to readily shift between ferrous ( $Fe^{2+}$ ) and ferric ( $Fe^{3+}$ ) oxidative states for redox catalysis or electron transport. This ability to readily accept and donate electrons also makes iron highly volatile, necessitating a well-coordinated iron transport and storage system in metazoans to prevent the production of toxic free radicals [4]. The sequestration of free iron by host proteins simultaneously prevents acquisition by microbes, a protective effect termed **nutritional immunity** (see *Glossary*) [5,6]. While appreciation has grown for the role of nutrient metals in infection, these 'battles for iron' and other trace metals provide intriguing cases for investigation from an evolutionary perspective. Here we discuss emerging questions on the control of iron in microbial infection and highlight recent and potential future insights regarding the evolution of molecular arms races, host range, microbial competition, and pathogen virulence.

### The Battle for Iron

A potential role for iron in immunity became apparent following an elegant series of experiments by Arthur Schade and Leona Caroline in the early 1940s [7]. While attempting to develop a vaccine against *Shigella*, the researchers observed that addition of raw egg white to their culture media severely inhibited the growth of diverse bacteria as well as fungi. The antiseptic properties of egg white have in fact been recognized since the days of Shakespeare, where it was applied to wounds during Act III of *King Lear*. While various nutrient supplements failed to reverse the

### Trends

The battle between microbes and their hosts for nutrient iron is emerging as a new front of evolutionary genetic conflict.

Molecular arms races can emerge between host iron-binding proteins and microbial 'iron piracy' factors that steal this nutrient for growth. Such rapid evolution may also contribute to the host range of pathogenic microbes.

Iron acquisition plays an important role in evolutionary interactions between microbes, both in the environment and within the host. Competition for iron can prevent infection by pathogens, while genetic changes in iron acquisition systems can enhance microbial virulence.

Evolutionary conflicts for nutrient iron are revealing potential new genetic mechanisms of disease resistance as well as avenues for therapeutic development.

<sup>1</sup>Department of Human Genetics, University of Utah School of Medicine, Salt Lake City, UT 84112, USA

\*Correspondence: [mbarber@genetics.utah.edu](mailto:mbarber@genetics.utah.edu) (M.F. Barber).

antimicrobial effect of egg whites, incinerated yeast extract did, suggesting that the limiting component was elemental in nature. Of 31 individual elements tested, supplementation with iron alone was sufficient to restore microbial growth in the presence of egg white. Adding to the fortuitous nature of their discovery, the authors posited that an iron-binding component present in the egg white prevented acquisition of this nutrient by microbes, which could have important implications for immunity. Two years later the scientists reported similar activity present in human blood serum [8]. The factor responsible for this activity in both cases was later revealed to be the protein **transferrin**, which plays a central role in animal iron metabolism by binding and transporting this metal to target cells [9,10].

In the decades following Schade and Caroline's initial discoveries, Eugene Weinberg proposed that withholding iron from microbial pathogens provided an important cornerstone of host defense, which he termed nutritional immunity [11]. Weinberg's theory explained previous observations that human iron overload disorders such as hereditary hemochromatosis and thalassemia render affected individuals highly susceptible to bacterial and fungal infections. The theory of nutritional immunity was also consistent with George Cartwright's earlier observations that infection induces an acute reduction in circulating iron levels [12–14]. Subsequent microbiology and molecular genetic studies established that nutritional immunity plays a pivotal role in defense against an array of pathogens, including bacteria, fungi, and parasites [3,15]. Owing to the iron-binding properties of proteins, such as transferrin, circulating levels of free iron in the body are orders of magnitude below the requirements for optimal microbial growth.

Microbes respond to iron starvation by actively scavenging this nutrient from host proteins to meet their metabolic requirements (Figure 1) [16]. One of the most common microbial iron acquisition strategies involves the secretion of **siderophores**, small molecule chelators, which possess an affinity for iron unmatched even by proteins such as transferrin [17,18]. Microbes then recover iron–siderophore complexes via cell surface receptors. Obviating the need for siderophores, several microbes also express receptors that directly recognize and extract iron from host proteins including transferrin and lactoferrin [19–23]. Additional mechanisms involve the acquisition of heme, the iron-containing porphyrin cofactor, from abundant host proteins such as hemoglobin [24–26]. Ferric reductases are an important class of iron acquisition systems in fungal pathogens, which convert transferrin or lactoferrin-bound ferric iron into a soluble ferrous form [27]. The identification of iron acquisition genes as pathogen **virulence factors** further underscores the role of iron in infection, as well as the potential for evolutionary conflicts to arise in the struggle for this precious nutrient.

### New Perspectives on Ancient Evolutionary Arms Races

Novel mutations that alter host–pathogen interactions can provide a substantial fitness advantage and spread in a population through **positive selection**. Recurrent bouts of positive selection at such interfaces can give rise to so-called 'molecular arms races', in which the host and pathogen must continually adapt to maintain comparative fitness [1]. Genes subject to such evolutionary conflicts are often characterized by an increased rate of nonsynonymous to synonymous substitutions (termed  $dN/dS$  or  $\omega$ ), reflecting recurrent selection for novel amino acid substitutions that alter protein interaction surfaces. Instances of such molecular arms races also exemplify Leigh Van Valen's **Red Queen Hypothesis**, which proposed that antagonistic coevolution leads to a perpetual cycle of adaptation in which neither opponent gains a permanent advantage [28]. Several core components of the vertebrate immune system have subsequently been shown to engage in such conflicts, some of which are able to dictate the outcome of an infection [29–35].

Our recent work highlighted the battle for iron as a new interface for Red Queen evolutionary conflicts [36]. As described earlier, transferrin was among the first vertebrate proteins to be

### Glossary

**Antagonistic pleiotropy:** also termed an evolutionary 'trade-off', in which a single gene controls multiple traits with opposing beneficial and deleterious effects.

**Black Queen Hypothesis:** describes the process by which gene loss can progress via natural selection, particularly in cases where individuals reduce investment in costly metabolic functions provided by other members of a microbial community.

**Microbiota:** the collection of microorganisms inhabiting a particular environment, such as the human body.

**Nutritional immunity:** a host immune defense mechanism by which essential nutrients, such as iron, are withheld in order to limit microbial growth and prevent infection.

**Positive selection:** the process by which new, beneficial genetic variation accrues in a population.

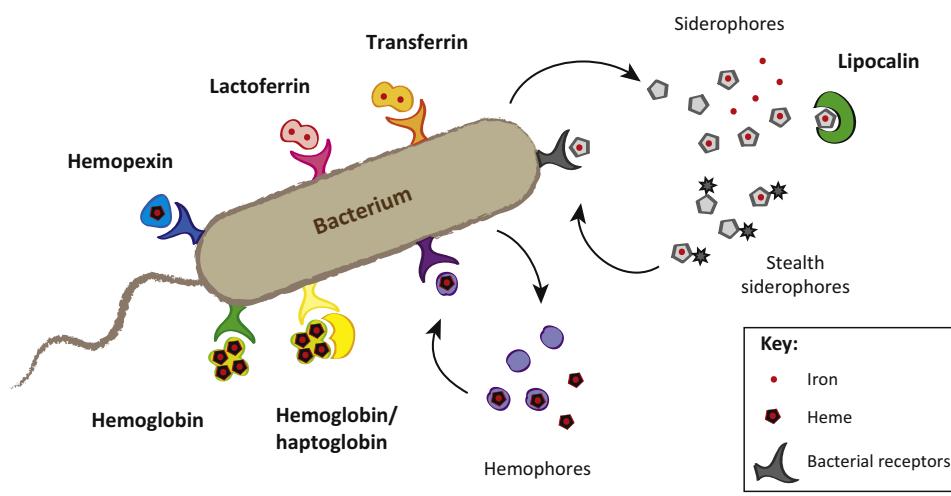
**Red Queen Hypothesis:** posits that antagonistic coevolution (e.g., between predators and prey or pathogens and hosts) produces a state in which constant adaptation is required to maintain comparative evolutionary fitness.

**Siderophore:** a diverse class of small molecule iron chelators, which are secreted by microbes and then internalized via surface receptors to mediate iron acquisition.

**Transferrin:** a serum glycoprotein in animals containing two iron-binding 'lobe' domains that deliver ferric iron to host cells via receptor-mediated endocytosis, as well as withholding iron from microbes.

**Virulence factor:** a gene or molecule that contributes to microbial infection, while not necessarily required for viability in nonpathogenic settings.

**Zoonosis:** an infectious disease naturally transmitted from animals to humans.

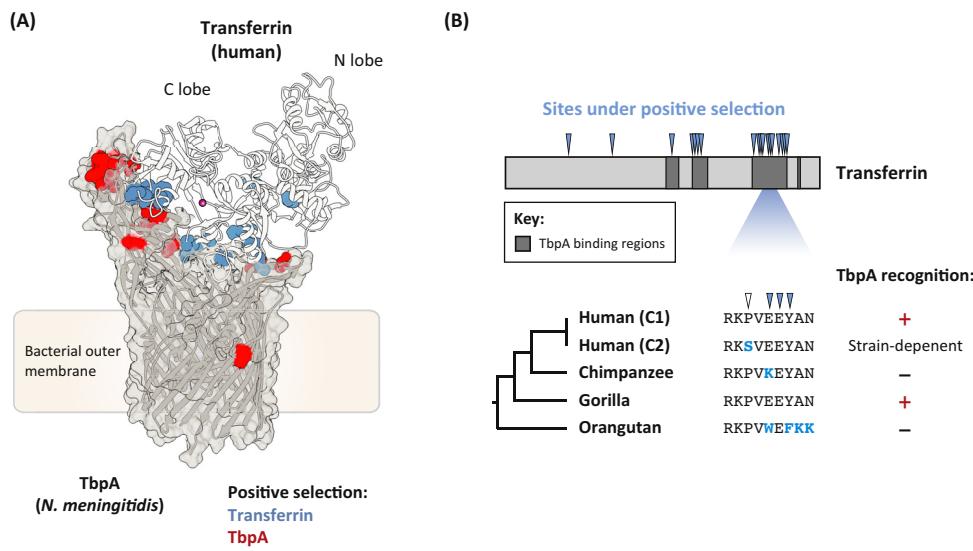


**Figure 1. Nutritional Immunity and Microbial Iron Piracy.** Illustration highlighting major components of bacterial iron acquisition, including surface receptors as well as secreted siderophores and hemophores. Host nutritional immunity proteins are denoted in bold.

implicated in nutritional immunity and is also a frequent target of iron acquisition by microbes. Reasoning that transferrin could be a focal point for genetic conflicts with pathogens, we performed phylogenetic analyses of transferrin gene divergence in the primate lineage. Not only has transferrin been subject to strong positive selection in primates but also rapidly evolving sites almost entirely overlap with the binding interface of a bacterial surface receptor, transferrin-binding protein A (TbpA), an important virulence factor in several human pathogens including *Neisseria meningitidis*, *Neisseria gonorrhoeae*, *Haemophilus influenzae*, and *Moraxella catarrhalis*, as well as a number of agricultural pathogens (Figure 2A) [21,37–41]. Single amino acid substitutions at rapidly evolving sites in transferrin were sufficient to control TbpA-binding specificities between related primates as well as for an abundant human transferrin variant, termed C2 (Figure 2B). Genetic signatures of positive selection at the transferrin–TbpA binding interface suggest that this interaction has been a key determinant of infection during millions of years of primate divergence. More broadly, these results demonstrate that nutritional immunity, similar to more established immune pathways, has strongly impacted host fitness during our long and intertwined history with microbes.

Evidence for an evolutionary arms race between transferrin and TbpA raises the question as to whether other host nutritional immunity factors may be subject to similar conflicts. Many pathogens encode receptors for other host iron-binding proteins including lactoferrin, a transferrin paralog expressed in milk, saliva, tears, mucus, and the secondary granules of neutrophils [37,42–44]. The evolution of lactoferrin introduces a fascinating twist; in addition to sequestering iron, lactoferrin has acquired mutations to generate antimicrobial peptide (AMP) domains that bind and disrupt pathogen membranes [45,46]. Many pathogens in turn encode factors that either scavenge lactoferrin-bound iron or inhibit associated AMP activity [47–49]. How these distinct functions have shaped lactoferrin evolution or potential arms races with pathogens remains to be determined.

Genetic conflicts in nutritional immunity may also unfold by means other than simple point mutation and selection at protein interaction sites. For example, the vertebrate protein lipocalin 2 (also known as siderocalin or NGAL) is a potent innate immunity factor that functions in part through binding and sequestration of siderophores, preventing their uptake by microbes [50,51].



**Figure 2. Evolutionary Conflict at the Transferrin–Transferrin-binding protein A (TbpA) interface.** (A) Cocrystal structure (Protein Data Bank: 3V8X) of human transferrin bound to TbpA from *Neisseria meningitidis*. Side chains of rapidly evolving amino acid positions in primate transferrin are shown in blue, with rapidly evolving TbpA sites among human pathogens shown in red (as described in [36]). (B) Schematic highlighting rapidly evolving regions in primate transferrin. Sites subject to positive selection are denoted with blue arrows; a variable site in humans (the C2 variant) is marked by a white arrow. Divergent amino acids among humans and other primates are shown in blue, and the ability of human-adapted TbpA to recognize each transferrin ortholog is shown on the right. The human transferrin C2 variant is recognized by TbpA from some but not all pathogens.

Some pathogens evade this defense through production of modified ‘stealth siderophores’, which are not recognized by lipocalin 2 [52,53]. Whether lipocalin 2 in turn has undergone adaptation resulting in enhanced or modified siderophore recognition is unknown. Understanding the extent to which molecular arms races have influenced other nutritional immunity factors beyond transferrin could reveal additional modes of adaptation underlying host–pathogen evolutionary conflicts.

The barrier imposed by nutritional immunity has seemingly produced an even more drastic evasion strategy by one pathogen – giving up iron altogether. Previous work has demonstrated that the bacterial spirochete *Borrelia burgdorferi*, the causative agent of Lyme disease, lacks a requirement for iron shared by nearly all other organisms [54]. This was an astounding discovery given that iron serves as a cofactor for numerous metalloproteins involved in essential cellular processes including the electron transport chain and DNA metabolism. How then has *B. burgdorferi* managed such an evolutionary feat? Closer inspection of the *B. burgdorferi* genome revealed that numerous genes encoding iron-binding proteins have been lost, and remaining enzymes that normally bind iron have undergone modification to bind manganese in its place [54,55]. Beyond these general observations, we are only beginning to unravel the stepwise genetic mechanisms that lead to such major evolutionary innovations [56–58]. Identifying other microbes that have foregone the requirement for iron could provide useful comparison points to understand the mechanics of complex evolutionary transitions.

Because many host nutritional immunity proteins also carry out crucial ‘day jobs’ in metal metabolism or transport, it is conceivable that **antagonistic pleiotropy**, or an evolutionary trade-off, could arise from an arms race with adverse consequences for the host. Sickle cell

anemia in humans provides a quintessential example, whereby hemoglobin mutations confer resistance to malaria infection at the expense of severe anemia in homozygous carriers [59]. Hereditary hemochromatosis (HH) is a condition characterized by increased iron absorption in the gut as well as serum iron overload, leading to iron accumulation in various organs and subsequent tissue damage [9,60]. HH caused by the C282Y mutation in the *HFE* gene is the most common genetic disorder among those of European descent, carried by approximately 10% of these individuals. Although the molecular mechanisms by which *HFE* mutations cause HH are still unclear, *HFE* is expressed on the surface of several cell types where it interacts with the transferrin receptor Tf-R to regulate iron absorption in the gut and release of iron stored in circulating macrophages. The high frequency of the C282Y mutation among Europeans has led to speculation as to the underlying cause for its abundance [61]. In addition to other associated health problems, individuals with HH are highly susceptible to infection by normally noninvasive microbes, such as the bacteria *Vibrio vulnificus* and *Yersinia enterocolitica* [62–64]. Ironically, this increased susceptibility to extracellular pathogens may be offset by resistance to others that normally infect macrophages such as *Mycobacterium tuberculosis* or *Salmonella enterica* serovar Typhi, which cause tuberculosis and typhoid fever, respectively [65]. While many questions remain regarding the consequences of HH mutations, these studies provide fascinating examples of how the role of iron in infection may contribute to instances of antagonistic pleiotropy in human genetic disorders.

### Iron in Host Range and Zoonoses

The term **zoonosis** refers to an infectious disease of animals that can be transmitted to humans. Because naïve populations typically lack pre-existing genetic resistance to these new pathogens, zoonotic diseases have caused some of the most deadly epidemics in human history, including the Black Death and the 1918 Spanish flu, along with recently emerging pathogens such as Ebola virus and the MERS coronavirus [66,67]. The mechanisms that dictate the host range of pathogens are thus of high interest to evolutionary biologists and infectious disease researchers alike. A number of studies have now implicated nutritional immunity factors in restricting the host range of several bacterial pathogens. For example, bacteria that utilize transferrin receptors possess extremely narrow host ranges, such as the human-specific *N. gonorrhoeae* and the porcine pathogen *Actinobacillus pleuropneumoniae*. Consistent with this observation, TbpA and its coreceptor TbpB exclusively recognize their host transferrin proteins when compared with diverse mammals [68–70]. Our recent work further demonstrated that single rapidly evolving sites in transferrin are sufficient to dictate TbpA-binding differences between even our closest relatives, such as chimpanzees [36]. Moreover, expressing or injecting human transferrin in mice can promote infection with human-specific *Neisseria* [71,72]. These findings suggest that adaptive evolution of nutritional immunity factors may be sufficient to establish a host range barrier against pathogens. The implications for transferrin variation on host range likely extend beyond bacteria as well, given that transferrin receptors have been implicated in iron acquisition and tropism of *Trypanosoma brucei*, the eukaryotic parasite that causes African sleeping sickness [73,74].

In addition to transferrin, it is likely that other nutritional immunity proteins contribute to limiting pathogen host range. Elegant work by Pishchany *et al.* demonstrated that the Gram-positive bacterium *Staphylococcus aureus* exhibits strong preference for human hemoglobin over mouse hemoglobin, which correlates with bacterial growth in murine models of infection [75]. While hemoglobin evolution has been extensively studied in the context of both environmental adaptation and malaria parasite resistance [59,76], the implications for this variation on nutritional immunity against bacterial pathogens has not been investigated. It is also notable that pathogens exhibiting restricted host iron requirements, including *N. gonorrhoeae* and *S. aureus*, pose urgent public health concerns given their increasing resistance to conventional antibiotics [77]. Applying genetic variants of transferrin to protein-based therapeutics provides one new means of treating infectious disease [78]. Therapeutic strategies building from evolutionary

studies of nutritional immunity could therefore provide new weapons against increasingly dire scenarios of resistance to traditional antimicrobial treatments.

### Microbial Competition for Iron: The Red Queen is Back in Black

The biology of competition has been a long-standing area of interest for both evolutionary theorists and microbial ecologists. A recent contribution to the field of microbial evolutionary theory came with the **Black Queen Hypothesis** [79]. The metaphor stems from the card game Hearts, in which players avoid holding the Queen of Spades or face a significant point penalty. The Black Queen Hypothesis posits that gene loss can be adaptive and proceed by natural selection, allowing individuals to reap public goods provided by other members of the microbial community. Iron acquisition is one such 'leaky' biological process that may be particularly prone to Black Queen conflicts. It has been widely observed that bacteria that do not produce siderophores nonetheless express siderophore receptors, allowing them to harvest this resource at the expense of their neighbors [16]. This hypothesis also invokes the long-standing concept of evolutionary 'cheaters', which can profoundly influence the stability of microbial populations. Previous studies have demonstrated that bacterial siderophore production follows many predictions of kin selection, whereby relatedness and degrees of competition influence the emergence of cheaters that do not produce siderophores [80,81]. Iron acquisition therefore provides as an informative system in which to study microbial population biology and evolution, including Black Queen dynamics.

The link between iron and microbial competition during infection was further illuminated in recent work by Deriu *et al.*, focusing on interactions between pathogenic *Salmonella* and commensal *Escherichia coli* in the gut [82]. The Nissle 1917 strain of *E. coli* was isolated from a soldier during World War I who appeared resistant to an outbreak of dysentery, and has subsequently been applied as a probiotic treatment for gastrointestinal ailments including ulcerative colitis and Crohn's disease [83,84]. The authors demonstrated that Nissle is able to suppress gastroenteritis induced by *Salmonella enterica* serovar Typhimurium through competition for iron [82]. The ability of Nissle to outcompete *S. enterica* was also dependent on the presence of the host siderophore-binding protein lipocalin 2, illustrating a complex interplay among host, pathogen, and commensal species. These findings further illustrate how microbial Black Queen dynamics can impact human disease, as well as the potential application of beneficial microbes to combat pathogens through iron sequestration.

The Red Queen and Black Queen Hypotheses highlight distinct modes of evolutionary adaptation ranging from antagonistic arms races to adaptive gene loss [85]. In the case of microbial iron piracy, both processes appear to play roles influencing fundamental functions. In addition to previous examples, loss of iron acquisition genes in *B. burgdorferi* could be interpreted as a Black Queen process arising within the host cell, whereas rapid evolution of bacterial transferrin receptors reflects a prototypical Red Queen conflict. The Black Queen Hypothesis may also provide a basis for understanding the fitness advantage conferred by bacterial receptors such as TbpA. By forgoing siderophore production and targeting host iron-binding proteins directly, these bacteria use a less leaky system, which may be inherently resistant to cheaters. In turn, dependence on these receptors gives rise to Red Queen conflicts with host proteins, such as transferrin, contributing to narrow host ranges observed for these strains. Future studies of iron acquisition could reveal additional genetic or ecological factors that contribute to these distinct evolutionary outcomes and the implications for infectious disease.

### Iron in the Evolution of Virulence

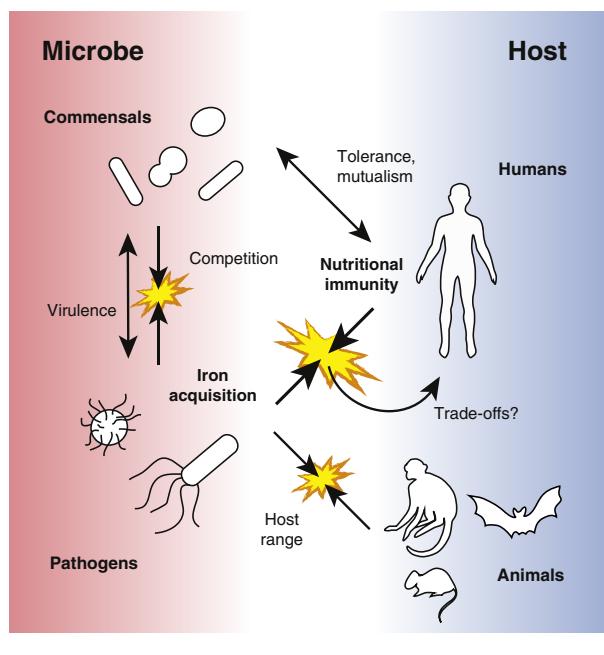
In addition to influencing interactions between microbes, iron can also regulate evolutionary transitions between commensal and pathogenic states. Iron acquisition genes are now

established as microbial virulence factors, in the sense that loss of these genes impairs pathogenicity without completely compromising organism viability [3,86]. However, a simple interpretation of iron acquisition dictating pathogenesis breaks down when considering the commensal **microbiota**. In nearly all cases these organisms require iron for survival, and yet rarely if ever cause disease. The role of iron in microbial virulence is thus more nuanced than it first appears and represents a growing area of research.

New insights on the role of iron in bacterial virulence were recently provided by studies of the opportunistic bacterial pathogen *Pseudomonas aeruginosa*. While *P. aeruginosa* is commonly found in the environment and is typically avirulent in immune-competent individuals, it readily colonizes the lungs of patients with cystic fibrosis (CF) where it poses a major source of mortality [87]. *P. aeruginosa*, like many microbes, possesses multiple distinct iron acquisition systems, including siderophores and a heme uptake system. Recently Marvig *et al.* investigated the genetic basis of host adaptation in *P. aeruginosa* strains that had maintained persistent human colonization for over 36 years [88]. Among several potential adaptations that took place during this period, evidence of positive selection was detected in the promoter of *phu*, the bacterial heme uptake system. Using experimental culture systems, the authors determined that increased expression of the *phu* system enhanced bacterial growth in a model of the CF lung, and that similar mutations had occurred independently in *P. aeruginosa* isolates from other patients with CF. These results suggest that a shift to enhanced heme uptake via hemoglobin plays an important role in the transition of *P. aeruginosa* from an environmental microbe to a dedicated human pathogen.

Iron acquisition can also dictate the ability of a microbe to shift between commensalism and virulence as conditions change, as was recently observed in the yeast *Candida albicans*. While often present as a commensal member of the gut microbiota, *C. albicans* is also capable of proliferating in the bloodstream and causing systemic infections in susceptible individuals [89]. The mechanisms by which *C. albicans* can shift between these distinct lifestyles are still largely mysterious. Chen *et al.* dissected the evolution of an elaborate transcriptional network in *C. albicans* whereby incorporation of a novel transcriptional activator, Sef1, into a pre-existing iron-responsive repressor system mediates growth in different host niches [90]. The authors demonstrated that Sef1 promotes iron acquisition and is essential for virulence *in vivo*. In addition, Sef1 is regulated by and itself regulates Sfu1, a transcriptional repressor. Sfu1 in turn represses iron acquisition and is dispensable for virulence, but is conversely essential for commensal growth in the gastrointestinal tract. Thus, *C. albicans* has evolved an intricate transcriptional program to regulate iron acquisition during commensal or pathogenic states [91]. Given that iron starvation acts as a trigger for activation of virulence genes in many pathogens [92], it is likely that similar gene regulatory networks also exist in other microbes.

Our understanding of iron in the control of commensal versus pathogenic states is only in its infancy. For example, while transferrin receptors are important virulence factors in pathogenic *N. meningitidis* and *N. gonorrhoeae*, these genes are also harbored by commensal *Neisseria* that inhabit the nasopharynx [93]. Adding to this complexity, *Neisseria* are naturally competent and horizontal gene transfer readily occurs between pathogenic and commensal strains [94]. Commensal microbes may therefore provide a genetic stockpile of virulence factors available for acquisition by pathogens. As we continue to learn more about the impact of the microbiota on human health and disease, these studies may also shed light on the role of iron in evolutionary interactions between seemingly beneficial or pathogenic inhabitants of our bodies.



Trends in Genetics

**Figure 3. Implications for Iron Piracy in Host–Microbe Evolution.** Overview depicting the roles of iron piracy and nutritional immunity in diverse evolutionary processes involving microbes and animal hosts.

### Outstanding Questions

In addition to transferrin, what other nutritional immunity factors are subject to molecular arms races with pathogens?

Do evolutionary conflicts exist for other nutrient metals such as manganese or zinc and their respective binding proteins?

How can we leverage evolution-guided studies of nutritional immunity to combat infectious disease?

### Concluding Remarks and Future Directions

Studies of microbial iron acquisition are proving a useful resource for investigating diverse evolutionary genetic phenomena (Figure 3). As new insights emerge, additional questions continue to arise (see Outstanding Questions). For example, while the majority of studies addressing nutritional immunity involve iron, recent work has begun to reveal the contribution of other essential metals including manganese and zinc in this process [95–98]. In addition, while previous findings have demonstrated that iron plays an important role in microbial competition, we understand comparatively little regarding its importance in the evolution of cooperative microbial communities. Finally, we have focused largely here on the role of bacteria in the evolutionary battle for iron, while fungi and parasites are undoubtedly also subject to similar conflicts [74,99,100]. The potential for adaptive evolution to dictate the outcome of microbial infection further provides an impetus to harness these studies for novel therapeutics. The application of evolutionary insights has led to promising genetic and chemical strategies for combatting human disease, most notably in the case of HIV [66,101–103]. Future studies promise to reveal additional genetic innovations favoring or countering microbial piracy of iron and other critical nutrients.

### Acknowledgments

We thank members of the Elde laboratory and Nicola Barber for helpful comments and suggestions on the manuscript. This work has been supported by awards from the National Institutes of Health to N.C.E. (R00GM090042) and M.F.B. (F32GM108288, K99GM115822). N.C.E. is a Pew Scholar in the Biomedical Sciences and the Mario R. Capecchi Endowed Chair in Genetics.

### References

1. Daugherty, M.D. and Malik, H.S. (2012) Rules of engagement: molecular insights from host–virus arms races. *Annu. Rev. Genet.* 46, 677–700
2. Palmer, A.C. and Kishony, R. (2013) Understanding, predicting and manipulating the genotypic evolution of antibiotic resistance. *Nat. Rev. Genet.* 14, 243–248
3. Cassat, J.E. and Skaar, E.P. (2013) Iron in infection and immunity. *Cell Host Microbe* 13, 509–519
4. Andrews, N.C. (1999) Disorders of iron metabolism. *N. Engl. J. Med.* 341, 1986–1995
5. Hood, M.I. and Skaar, E.P. (2012) Nutritional immunity: transition metals at the pathogen–host interface. *Nat. Rev. Microbiol.* 10, 525–537
6. Potrykus, J. et al. (2014) Conflicting interests in the pathogen–host tug of war: fungal micronutrient scavenging versus mammalian nutritional immunity. *PLoS Pathog.* 10, e1003910
7. Schade, A.L. and Caroline, L. (1944) Raw hen egg white and the role of iron in growth inhibition of *Shigella dysenteriae*, *Staphylococcus aureus*, *Escherichia coli* and *Saccharomyces cerevisiae*. *Science* 100, 14–15

8. Schade, A.L. and Caroline, L. (1946) An iron-binding component in human blood plasma. *Science* 104, 340–341
9. Ganz, T. and Nemeth, E. (2011) Hepcidin and disorders of iron metabolism. *Annu. Rev. Med.* 62, 347–360
10. Gkouvatsos, K. *et al.* (2012) Regulation of iron transport and the role of transferrin. *Biochim. Biophys. Acta* 1820, 188–202
11. Weinberg, E.D. (1975) Nutritional immunity. Host's attempt to withhold iron from microbial invaders. *JAMA* 231, 39–41
12. Weinberg, E.D. (1978) Iron and infection. *Microbiol. Rev.* 42, 45–66
13. Weinberg, E.D. (1984) Iron withholding: a defense against infection and neoplasia. *Physiol. Rev.* 64, 65–102
14. Cartwright, G.E. *et al.* (1946) The anemia associated with chronic infection. *Science* 103, 72–73
15. Skaar, E.P. (2010) The battle for iron between bacterial pathogens and their vertebrate hosts. *PLoS Pathog.* 6, e1000949
16. Rattleidge, C. and Dover, L.G. (2000) Iron metabolism in pathogenic bacteria. *Annu. Rev. Microbiol.* 51, 881–941
17. Francis, J. *et al.* (1953) Mycobactin, a growth factor for *Mycobacterium johnei*. I. Isolation from *Mycobacterium phlei*. *Biochem. J.* 55, 596–607
18. Holden, V.I. and Bachman, M.A. (2015) Diverging roles of bacterial siderophores during infection. *Metallomics* 7, 986–995
19. Schryvers, A.B. and Morris, L.J. (1988) Identification and characterization of the human lactoferrin-binding protein from *Neisseria meningitidis*. *Infect. Immun.* 56, 1144–1149
20. Lee, B.C. and Schryvers, A.B. (1988) Specificity of the lactoferrin and transferrin receptors in *Neisseria gonorrhoeae*. *Mol. Microbiol.* 2, 827–829
21. Cornelissen, C.N. *et al.* (1992) Gonococcal transferrin-binding protein 1 is required for transferrin utilization and is homologous to TonB-dependent outer membrane receptors. *J. Bacteriol.* 174, 5788–5797
22. Salmon, D. *et al.* (1994) A novel heterodimeric transferrin receptor encoded by a pair of VSG expression site-associated genes in *T. brucei*. *Cell* 78, 75–86
23. Steverding, D. (2000) The transferrin receptor of *Trypanosoma brucei*. *Parasitol. Int.* 48, 191–198
24. Lewis, L.A. and Dyer, D.W. (1995) Identification of an iron-regulated outer membrane protein of *Neisseria meningitidis* involved in the utilization of hemoglobin complexed to haptoglobin. *J. Bacteriol.* 177, 1299–1306
25. Torres, V.J. *et al.* (2006) *Staphylococcus aureus* IsdB is a hemoglobin receptor required for heme iron utilization. *J. Bacteriol.* 188, 8421–8429
26. Fabian, M. *et al.* (2009) Heme transfer to the bacterial cell envelope occurs via a secreted hemophore in the Gram-positive pathogen *Bacillus anthracis*. *J. Biol. Chem.* 284, 32138–32146
27. Saikia, S. *et al.* (2014) Role of ferric reductases in iron acquisition and virulence in the fungal pathogen *Cryptococcus neoformans*. *Infect. Immun.* 82, 839–850
28. Van Valen, L. (1973) A new evolutionary law. *Evol. Theory* 1, 1–30
29. Sawyer, S.L. *et al.* (2004) Ancient adaptive evolution of the primate antiviral DNA-editing enzyme APOBEC3G. *PLoS Biol.* 2, e275
30. Sawyer, S.L. *et al.* (2005) Positive selection of primate TRIM5 $\alpha$  identifies a critical species-specific retroviral restriction domain. *Proc. Natl. Acad. Sci. U.S.A.* 102, 2832–2837
31. Elde, N.C. *et al.* (2008) Protein kinase R reveals an evolutionary model for defeating viral mimicry. *Nature* 457, 485–489
32. Mitchell, P.S. *et al.* (2012) Evolution-guided identification of anti-viral specificity determinants in the broadly acting interferon-induced innate immunity factor MxA. *Cell Host Microbe* 12, 598–604
33. Patel, M.R. *et al.* (2012) Convergent evolution of escape from hepcivirus antagonism in primates. *PLoS Biol.* 10, e1001282
34. Demogines, A. *et al.* (2013) Dual host-virus arms races shape an essential housekeeping protein. *PLoS Biol.* 11, e1001571
35. Sironi, M. *et al.* (2015) Evolutionary insights into host-pathogen interactions from mammalian sequence data. *Nat. Rev. Genet.* 16, 224–236
36. Barber, M.F. and Elde, N.C. (2014) Escape from bacterial iron piracy through rapid evolution of transferrin. *Science* 346, 1362–1366
37. Schryvers, A.B. (1988) Characterization of the human transferrin and lactoferrin receptors in *Haemophilus influenzae*. *Mol. Microbiol.* 2, 467–472
38. Cornelissen, C.N. *et al.* (1998) The transferrin receptor expressed by gonococcal strain FA1090 is required for the experimental infection of human male volunteers. *Mol. Microbiol.* 27, 611–616
39. Yu, R.H. and Schryvers, A.B. (1993) The interaction between human transferrin and transferrin binding protein 2 from *Moraxella (Branhamella) catarrhalis* differs from that of other human pathogens. *Microb. Pathog.* 15, 433–445
40. Moraes, T.F. *et al.* (2009) Insights into the bacterial transferrin receptor: the structure of transferrin-binding protein B from *Actinobacillus pleuropneumoniae*. *Mol. Cell* 35, 523–533
41. Noinaj, N. *et al.* (2012) Structural basis for iron piracy by pathogenic *Neisseria*. *Nature* 483, 53–58
42. García-Montoya, I.A. *et al.* (2012) Lactoferrin a multiple bioactive protein: an overview. *Biochim. Biophys. Acta* 1820, 226–236
43. Beddek, A.J. and Schryvers, A.B. (2010) The lactoferrin receptor complex in Gram negative bacteria. *Biometals* 23, 377–386
44. Noinaj, N. *et al.* (2013) Structural insight into the lactoferrin receptors from pathogenic *Neisseria*. *J. Struct. Biol.* 184, 83–92
45. Yamauchi, K. *et al.* (1993) Antibacterial activity of lactoferrin and a pepsin-derived lactoferrin peptide fragment. *Infect. Immun.* 61, 719–728
46. Haney, E.F. *et al.* (2009) Novel lactoferrapin antimicrobial peptides derived from human lactoferrin. *Biochimie* 91, 141–154
47. Hammerschmidt, S. *et al.* (1999) Identification of pneumococcal surface protein A as a lactoferrin-binding protein of *Streptococcus pneumoniae*. *Infect. Immun.* 67, 1683–1687
48. Senkovich, O. *et al.* (2007) Structure of a complex of human lactoferrin N-lobe with pneumococcal surface protein A provides insight into microbial defense mechanism. *J. Mol. Biol.* 370, 701–713
49. Deka, R.K. *et al.* (2007) Crystal structure of the Tp34 (TP0971) lipoprotein of *Treponema pallidum*: implications of its metal-bound state and affinity for human lactoferrin. *J. Biol. Chem.* 282, 5944–5958
50. Flo, T.H. *et al.* (2004) Lipocalin 2 mediates an innate immune response to bacterial infection by sequestrating iron. *Nature* 432, 917–921
51. Bachman, M.A. *et al.* (2012) Interaction of lipocalin 2, transferrin, and siderophores determines the replicative niche of *Klebsiella pneumoniae* during pneumonia. *MBio* 3, e00224–e311
52. Hantke, K. *et al.* (2003) Salmochelins, siderophores of *Salmonella enterica* and uropathogenic *Escherichia coli* strains, are recognized by the outer membrane receptor IroN. *Proc. Natl. Acad. Sci. U.S.A.* 100, 3677–3682
53. Abergel, R.J. *et al.* (2006) Anthrax pathogen evades the mammalian immune system through stealth siderophore production. *Proc. Natl. Acad. Sci. U.S.A.* 103, 18499–18503
54. Posey, J.E. and Gherardin, F.C. (2000) Lack of a role for iron in the Lyme disease pathogen. *Science* 288, 1651–1653
55. Aguirre, J.D. *et al.* (2013) A manganese-rich environment supports superoxide dismutase activity in a Lyme disease pathogen, *Borrelia burgdorferi*. *J. Biol. Chem.* 288, 8468–8478
56. Blount, Z.D. *et al.* (2012) Genomic analysis of a key innovation in an experimental *Escherichia coli* population. *Nature* 489, 513–518
57. Leiby, N. and Marx, C.J. (2014) Metabolic erosion primarily through mutation accumulation, and not tradeoffs, drives limited evolution of substrate specificity in *Escherichia coli*. *PLoS Biol.* 12, e1001789
58. Chou, H.H. *et al.* (2009) Fast growth increases the selective advantage of a mutation arising recurrently during evolution under metal limitation. *PLoS Genet.* 5, e1000652
59. Ashley-Koch, A. *et al.* (2000) Sickle hemoglobin (HbS) allele and sickle cell disease: a HuGE review. *Am. J. Epidemiol.* 151, 839–845

60. Vujić, M. (2014) Molecular basis of HFE-hemochromatosis. *Front. Pharmacol.* 5, 42
61. Weinberg, E.D. (2008) Survival advantage of the hemochromatosis C282Y mutation. *Perspect. Biol. Med.* 51, 98–102
62. Wright, A.C. *et al.* (1981) Role of iron in the pathogenesis of *Vibrio vulnificus* infections. *Infect. Immun.* 34, 503–507
63. Quenée, L.E. *et al.* (2012) Hereditary hemochromatosis restores the virulence of plague vaccine strains. *J. Infect. Dis.* 206, 1050–1058
64. Weinberg, E.D. (2000) Microbial pathogens with impaired ability to acquire host iron. *Biometals* 13, 85–89
65. Olakanmi, O. *et al.* (2007) Hereditary hemochromatosis results in decreased iron acquisition and growth by *Mycobacterium tuberculosis* within human macrophages. *J. Leukoc. Biol.* 81, 195–204
66. Sawyer, S.L. and Elde, N.C. (2012) A cross-species view on viruses. *Curr. Opin. Virol.* 2, 561–568
67. Karlsson, E.K. *et al.* (2014) Natural selection and infectious disease in human populations. *Nat. Rev. Genet.* 15, 379–393
68. Schryvers, A.B. and Gonzalez, G.C. (1990) Receptors for transferrin in pathogenic bacteria are specific for the host's protein. *Can. J. Microbiol.* 36, 145–147
69. Cornelissen, C.N. *et al.* (1993) Expression of gonococcal transferrin-binding protein 1 causes *Escherichia coli* to bind human transferrin. *J. Bacteriol.* 175, 2448–2450
70. Litt, D.J. *et al.* (2000) *Neisseria meningitidis* expressing transferrin binding proteins of *Actinobacillus pleuropneumoniae* can utilize porcine transferrin for growth. *Infect. Immun.* 68, 550–557
71. Oftung, F. *et al.* (1999) A mouse model utilising human transferrin to study protection against *Neisseria meningitidis* serogroup B induced by outer membrane vesicle vaccination. *FEMS Immunol. Med. Microbiol.* 26, 75–82
72. Zarantonelli, M.L. *et al.* (2007) Transgenic mice expressing human transferrin as a model for meningococcal infection. *Infect. Immun.* 75, 5609–5614
73. Bitter, W. *et al.* (1998) The role of transferrin-receptor variation in the host range of *Trypanosoma brucei*. *Nature* 491, 499–502
74. Steverding, D. (2003) The significance of transferrin receptor variation in *Trypanosoma brucei*. *Trends Parasitol.* 19, 125–127
75. Pishchany, G. *et al.* (2010) Specificity for human hemoglobin enhances *Staphylococcus aureus* infection. *Cell Host Microbe* 8, 544–550
76. Natarajan, C. *et al.* (2013) Epistasis among adaptive mutations in deer mouse hemoglobin. *Science* 340, 1324–1327
77. Blair, J.M.A. *et al.* (2015) Molecular mechanisms of antibiotic resistance. *Nat. Rev. Microbiol.* 13, 42–51
78. Lin, L. *et al.* (2014) Transferrin iron starvation therapy for lethal bacterial and fungal infections. *J. Infect. Dis.* 210, 254–264
79. Morris, J.J. *et al.* (2012) The Black Queen Hypothesis: evolution of dependencies through adaptive gene loss. *MBio* 3, e00036–12
80. Griffin, A.S. *et al.* (2004) Cooperation and competition in pathogenic bacteria. *Nature* 430, 1024–1027
81. Ghoul, M. *et al.* (2014) An experimental test of whether cheating is context dependent. *J. Evol. Biol.* 27, 551–556
82. Deriu, E. *et al.* (2013) Probiotic bacteria reduce *Salmonella typhimurium* intestinal colonization by competing for iron. *Cell Host Microbe* 14, 26–37
83. Nissle, A. (1959) [Explanations of the significance of colonic dysbacteria and the mechanism of action of *E. coli* therapy (mutaflor)]. *Medizinische* 4, 1017–1022 (in German)
84. Jacobi, C.A. and Malfertheiner, P. (2011) *Escherichia coli* Nissle 1917 (Mutaflor): new insights into an old probiotic bacterium. *Dig. Dis.* 29, 600–607
85. Morris, J.J. (2015) Black Queen evolution: the role of leakiness in structuring microbial communities. *Trends Genet.* 31, 475–482
86. Subashchandrabose, S. and Moleby, H.L.T. (2015) Back to the metal age: battle for metals at the host-pathogen interface during urinary tract infection. *Metalomics* 7, 935–942
87. Gellatly, S.L. and Hancock, R.E.W. (2013) *Pseudomonas aeruginosa*: new insights into pathogenesis and host defenses. *Pathog. Dis.* 67, 159–173
88. Marvig, R.L. *et al.* (2014) Within-host evolution of *Pseudomonas aeruginosa* reveals adaptation toward iron acquisition from hemoglobin. *MBio* 5, e00966–e1014
89. Cheng, S-C. *et al.* (2012) Interplay between *Candida albicans* and the mammalian innate host defense. *Infect. Immun.* 80, 1304–1313
90. Chen, C. *et al.* (2011) An iron homeostasis regulatory circuit with reciprocal roles in *Candida albicans* commensalism and pathogenesis. *Cell Host Microbe* 10, 118–135
91. Noble, S.M. (2013) *Candida albicans* specializations for iron homeostasis: from commensalism to virulence. *Curr. Opin. Microbiol.* 16, 708–715
92. Troxell, B. and Hassan, H.M. (2013) Transcriptional regulation by Ferric Uptake Regulator (Fur) in pathogenic bacteria. *Front. Cell Infect. Microbiol.* 3, 59
93. Marri, P.R. *et al.* (2010) Genome sequencing reveals widespread virulence gene exchange among human *Neisseria* species. *PLoS ONE* 5, e11835
94. Rotman, E. and Seifert, H.S. (2014) The genetics of *Neisseria* species. *Annu. Rev. Genet.* 48, 405–431
95. Liu, J.Z. *et al.* (2012) Zinc sequestration by the neutrophil protein calprotectin enhances *Salmonella* growth in the inflamed gut. *Cell Host Microbe* 11, 227–239
96. Gaddy, J.A. *et al.* (2014) The host protein calprotectin modulates the *Helicobacter pylori* cag type IV secretion system via zinc sequestration. *PLoS Pathog.* 10, e1004450
97. Corbin, B.D. *et al.* (2008) Metal chelation and inhibition of bacterial growth in tissue abscesses. *Science* 319, 962–965
98. Kehl-Fie, T.E. and Saar, E.P. (2010) Nutritional immunity beyond iron: a role for manganese and zinc. *Curr. Opin. Chem. Biol.* 14, 218–224
99. Kronstad, J.W. *et al.* (2013) An encapsulation of iron homeostasis and virulence in *Cryptococcus neoformans*. *Trends Microbiol.* 21, 457–465
100. Jung, W.H. and Do, E. (2013) Iron acquisition in the human fungal pathogen *Cryptococcus neoformans*. *Curr. Opin. Microbiol.* 16, 686–691
101. Richardson, M.W. *et al.* (2014) Stabilized human TRIM5 $\alpha$  protects human T cells from HIV-1 infection. *Mol. Ther.* 22, 1084–1095
102. Tebas, P. *et al.* (2014) Gene editing of CCR5 in autologous CD4 T cells of persons infected with HIV. *N. Engl. J. Med.* 370, 901–910
103. Van Der Ryst, E. (2015) Maraviroc – a CCR5 antagonist for the treatment of HIV-1 infection. *Front. Immunol.* 6, 277